

# HAEMATOLOGICAL AND SERUM BIOCHEMICAL INDICES OF STARTER BROILERS FED NEEM (*Azadirachta indica*) LEAF MEAL

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**ABSTRACT:** A 28-day feeding trial was conducted to evaluate the effects of dietary inclusion of Neem (*Azadirachta indica*) leaf meal on the haematological and serum biochemical indices of starter broilers. The Neem leaves used in the experiment were manually harvested, air-dried and milled to become Neem leaf meal. The Neem leaf meal was included in broiler starter diets at 0, 2.5, 5.0, 7.5 and 10% levels, respectively. One hundred and fifty (150) Anak broiler starter chicks raised on a commercial starter mash for one week were used. They were divided into 5 groups of 30 birds each and randomly assigned to the 5 experimental diets in a completely randomized design (CRD). Each group was sub-divided into 3 replicates of 10 birds each and each replicate housed in a pen fitted with necessary brooding facilities. Feed and water were given to them *ad libitum* for 4 weeks. Proximate analysis of the Neem leaf meal displayed same characteristics as leaf meals from other tropical browse plants - high crude fibre (15.56%) and moderate crude protein content (18.10%). At the end of the feeding trial, blood was collected from the birds, 4 per treatment and analysed for haematological and serum biochemical indices. Haemoglobin (Hb) and packed cell volume (PCV) of the birds were significantly reduced ( $P<0.05$ ) but not below the level considered normal for birds. No traces of monocytes, eosinophils and basophils were observed. Blood sugar was significantly raised ( $P<0.05$ ) by the leaf meal but cholesterol was significantly ( $P<0.05$ ) decreased. Alkaline phosphatase (ALP), alanine transaminase (ALT) and aspartate transaminase (AST) decreased with increase in leaf meal ( $P<0.05$ ). Serum electrolytes: calcium, sodium, potassium, chloride and bicarbonate tended to show that Neem leaf meal up to 10% dietary inclusion level could still maintain the integrity of the kidney in boosting cation/anion exchange. The haematological and serum biochemical parameters obtained from this study suggested that dietary Neem leaf meal has no deleterious effects on some physiological indices of starter broilers.

**Keywords:** Neem leaf meal, starter broilers, haematological, serum, biochemical indices.

## INTRODUCTION

The growth of human and livestock population which has created increased demand for food and feed in the developing countries suggests that alternative feed resources must be identified and evaluated (Nworgu et al., 2007). In evaluating such unconventional feed resources, it is important to also check the effects of such feed resources on the health status of the livestock. Esonu et al. (2001) stated that haematological constituents reflect the physiological responsiveness of the animals and the influence of diet on haematological traits is very strong (Church et al., 1984; Babatunde et al., 1987). Restricted low energy feed intake results in elevated MCHC. Haemoglobin and packed cell volume are very sensitive to the levels of protein intake as the values increase with increase in dietary protein concentration (Edoziem and Switzer, 1977). It has also been observed that serum urea, total protein and creatinine contents depend on both the quality and quantity of protein supplied in the diet (Iyayi and Tewe, 1998).

The use of leaf meals of plants as feed ingredients as alternative to conventional feed resources is a novel area of research in animal nutrition. Leaf meals of some tropical legumes and browse plants, rich in nutrients like vitamins, minerals and carotenoids have been reported (Vohra et al., 1972; Udedibie, 1987; Udedibie and Opara, 1996; Esonu et al., 2002).

One of the tropical plants that have attracted attention of animal nutritionists in recent time is the neem tree (*A. indica*). Various parts of the tree have medicinal value (Chakraborty et al., 1989) and recent studies by Esonu et al.

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(2006) have shown that its leaf meal could be of some value in the diet of laying hens both as feed ingredient and egg yolk pigments. There is the need to also evaluate its effects on haematological and serum biochemical constituents of poultry.

The studies herein reported were therefore designed to examine the haematological and serum biochemical indices of starter broilers as affected by dietary Neem leaf meal.

## MATERIALS AND METHODS

### Study sites

This study was carried out in the Poultry Unit of the Teaching and Research Farm of the School of Agriculture and Agricultural Technology and Animal Science Laboratory of the Federal University of Technology, Owerri, Imo State, Nigeria. Imo State lies between latitude 4° 4' and 6° 3' N and longitude 6° 15' and 8° 15' E. Owerri is about 100m above sea level. The climatic data of Owerri as summarized in Ministry of Lands and Survey Atlas (1984) of Imo State is as follows: mean annual rainfall, 2500 mm; temperature range, 26.5 – 27.5 °C and humidity range of 70 – 80 %. Dry season duration (i.e. months with less than 65mm rainfall) is 3months. The annual evapo-transpiration is 1450 mm and the soil type is essentially sandy loam with average pH of 5.5.

### Source and processing of Neem leaves

Fresh green Neem leaves used for the experiment were harvested within the University environment. Each batch of collection was air-dried. They were considered adequately dried when they became crispy to the touch. They were then milled, using a hammer mill with 2 mm sieve, to produce Neem leaf meal (NLM). Samples of the leaf meal were subjected to proximate analysis according to AOAC (1995).

### Experimental Diets

Five white maize-based experimental broiler starter diets (23% CP) were made, incorporating the leaf meal at 5 levels of 0.00, 2.50, 5.00, 7.50 and 10% designated as T<sub>0</sub>, T<sub>2.50</sub>, T<sub>5.0</sub>, T<sub>7.5</sub> and T<sub>10</sub>, respectively. The ingredient composition of the experimental diets is shown in Table 1. The diets were balanced for crude protein and caloric content as per the requirements of this class of birds in the tropics (Sansbury, 1980).

<b>Table 1 - Ingredient composition of the broiler starter experimental diets</b>					
<b>Ingredients (%)</b>	<b>Dietary levels of leaf meal (%)</b>				
	<b>0.00</b>	<b>2.50</b>	<b>5.00</b>	<b>7.50</b>	<b>10.00</b>
White Maize	50.00	49.00	47.00	46.00	45.00
Neem Leaf meal	0.00	2.50	5.00	7.50	10.00
Soybean meal	26.00	26.00	26.00	26.00	26.00
Wheat offal	10.00	8.50	8.00	6.50	5.00
Palm kernel cake	5.00	5.00	5.00	5.00	5.00
Fish meal	2.00	2.00	2.00	2.00	2.00
Blood meal	3.00	3.00	3.00	3.00	3.00
Bone meal	3.00	3.00	3.00	3.00	3.00
Common Salt	0.25	0.25	0.25	0.25	0.25
Vitamin/Trace min. premix	0.25	0.25	0.25	0.25	0.25
L-lysine	0.25	0.25	0.25	0.25	0.25
L-methionine	0.25	0.25	0.25	0.25	0.25
<b>Total</b>	<b>100.00</b>	<b>100.00</b>	<b>100.00</b>	<b>100.00</b>	<b>100.00</b>
<b>Calculated Analysis (% of Dm)</b>					
Crude protein	21.97	21.96	21.95	21.92	21.94
Crude fibre	4.41	4.70	5.00	5.28	5.57
Ash	4.02	4.06	4.11	4.16	4.20
Ether extract	4.34	4.32	4.33	4.31	4.30
Calcium	1.82	1.82	1.82	1.82	1.82
Phosphorus	0.98	0.98	0.97	0.96	0.95
Metabolizable Energy(kcal/kg)	2734.71	2702.91	2690.27	2683.45	2685.45
Each kg of feed contained: Vit. A, 2, 000,000 i.u.; vit. D <sub>3</sub> , 100 iu ; vit. E, 8g ; vit. K, 0.4g ; vit. B <sub>1</sub> , 0.3g ; vit. B <sub>2</sub> , 1.0g ; vit. B <sub>6</sub> , 0.6g ; vit. C, 2.40g, vit. B <sub>12</sub> , 40g ; Mn, 160g ; Fe, 8.0g ; Zn, 7.2g ; Cu, 0.3g ; Iodine, 0.25g ; Co, 36.0mg ; Se, 16.0mg.					

### Experimental Birds and Design

One hundred and fifty (150) Anak broiler chicks raised on a commercial starter mash for one week were used. They were divided into 5 groups of 30 birds each and each group randomly assigned to one of the 5 experimental diets in a completely randomized design (CRD). Each group was further sub-divided into 3 replicates of 10 birds each and each replicate housed in a pen fitted with necessary brooding facilities. Feed and water were given to them *ad libitum*. The birds were weighed at the beginning of the trial and thereafter, weekly. Daily feed intake per group was determined by weighing the feed offered and the left-over the following morning. The feeding trial lasted 4 weeks.

### Blood collection and Analysis

At the end of the feeding trial (5<sup>th</sup> week) blood samples were collected from one broiler bird per replicate, making three samples per treatment. Bleeding was done from the punctured wing vein with a 5 ml scalp vein needle set. About 2 ml of blood was collected from each bird into two sets of sterilized bottles, one containing ethylenediaminetetraacetic acid (EDTA) as the anti-coagulant for determination of haematological parameters. Haemoglobin concentration (Hb) was determined, using Sahli method and the value recorded in g/100 mls (WHO, 1980), RBC and WBC using the improved Neubauer haemocytometer as described by Dacie and Lewis (1991). PCV was determined by the Microhaematocrit method, while mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH) and mean corpuscular haemoglobin concentration (MCHC) were calculated using the appropriate formulae.

The second set of bottles without EDTA was centrifuged in a macro centrifuge to obtain serum for biochemical analysis. Total protein was determined, using the burette method as described by Dumas (1975); urea by dimethyl methyl monoxide method as described by Varley et al. (1980). Creatinine was determined by Jaffe reaction method as described by Henry et al. (1974). Albumin was measured using dye-binding technique with bromocresol green as described by Dumas and Biggs (1972). Serum potassium (K<sup>+</sup>) and sodium (Na<sup>+</sup>) were determined by the calorimetric method, while serum cholesterol was by a modification of the Liebermann- Burchard test.

### Statistical analysis

Data collected were subjected to analysis of variance (ANOVA). Where analysis of variance indicated significant treatment effects, the means were separated using Duncan's New Multiple Range Test as described by Steel and Torrie (1980).

### RESULTS AND DISCUSSION:

Proximate composition of the leaf meal is presented in Table 2. The leaf meal displayed same characteristics as leaf meals from other tropical browse plants – high crude fibre and moderate crude protein content as reported for *Jacaranda mimosifolia* (Okorie, 2006) and for *Microdesmis puberula* (Esonu et al., 2002). With relatively high crude fibre content (15.56%), the metabolizable energy must be low even though its gross energy content was high (4.16 Kcal/g).

Components	% of dm
Crude Protein	18.10
Crude Fibre	15.56
Ether Extract	2.50
Ash	5.62
Nitrogen free Extract	58.22
Gross Energy (Kcal/gm)	4.16

### Haematological Indices

The haematological indices of the starter broilers fed graded levels of Neem leaf meal are presented in table 3. The Hb value for T<sub>5.0</sub> compared favourably (P>0.05) with that of control. There were no significant differences (P>0.05) among the Hb values for T<sub>2.5</sub>, T<sub>7.5</sub> and T<sub>10</sub>. The values of Hb recorded by all the groups were, however, within the normal range for chicken (Aiello and Mays, 1998; Okeudo, 2003). There was significant difference (P<0.05) between the ESR values for T<sub>0</sub> and those of T<sub>7.5</sub> and T<sub>10</sub>. There were no significant differences (P>0.05) among the control and those of T<sub>2.5</sub> and T<sub>5.0</sub>. The ESR was rather decreasing with increase in leaf meal inclusion. Sedimentation rates are increased in cases of acute general infection, malignant tumours and pregnancy.

**Table 3 - Effects of graded levels of Neem leaf meals on the haematological parameters of starter broilers**

Indices	Dietary levels of Neem leaf meal					SEM
	T <sub>0</sub>	T <sub>2.5</sub>	T <sub>5.0</sub>	T <sub>7.5</sub>	T <sub>10</sub>	
Hb (g/dl)	9.60 <sup>a</sup>	8.00 <sup>b</sup>	9.17 <sup>ab</sup>	8.40 <sup>b</sup>	8.67 <sup>b</sup>	0.28
ESR (Mm/1 <sup>hr</sup> )	6.33 <sup>a</sup>	4.67 <sup>a</sup>	4.67 <sup>a</sup>	2.00 <sup>b</sup>	3.00 <sup>b</sup>	0.75
TWBC x 10 <sup>3</sup> (uL)	1.143 <sup>a</sup>	0.787 <sup>b</sup>	1.325 <sup>a</sup>	1.578 <sup>a</sup>	1.119 <sup>a</sup>	0.08
PCV (%)	28.33 <sup>a</sup>	23.33 <sup>b</sup>	27.33 <sup>a</sup>	25.00 <sup>b</sup>	26.00 <sup>ab</sup>	0.87
Heterophils (%)	4.00 <sup>b</sup>	4.67 <sup>b</sup>	5.00 <sup>b</sup>	6.00 <sup>a</sup>	4.00 <sup>b</sup>	0.45
Lymphocytes (%)	96.00 <sup>a</sup>	95.33 <sup>a</sup>	95.00 <sup>a</sup>	95.33 <sup>a</sup>	96.00 <sup>a</sup>	0.45
Monocytes (%)	0.00	0.00	0.00	0.00	0.00	0.00
Eosinophils (%)	0.00	0.00	0.00	0.00	0.00	0.00
Basophils (%)	0.00	0.00	0.00	0.00	0.00	0.00

<sup>ab</sup>Means in the same row with different superscripts are significantly different (P < 0.05)

It shows therefore that the birds did not suffer from any of the aforementioned. Except for T<sub>2.5</sub>, the values of TWBC for the Neem leaf groups compared favourably with that of the control and also were within the normal range. There were significant differences (P<0.05) between the PCV of the control and those of T<sub>2.5</sub> and T<sub>10</sub>. However, T<sub>5.0</sub> compared favourably (P>0.05) with the control. The lymphocytes were not significantly affected by the treatments (P>0.05). No traces of monocytes, eosinophils and basophils were observed in all the treatment groups as earlier reported by Akpan (2007). It therefore shows that Neem leaf meal did not produce any form of infection since these parameters only observed when there is infection (Frandsen, 1974).

### Serum Biochemical Indices

The serum biochemical constituents of the birds are shown in table 4. Neem leaf meal tended to elevate the blood glucose level of the birds while reducing the cholesterol level. The increase in blood sugar level as the dietary Neem leaf meal increased was quite interesting because birds generally maintain a high and relatively constant blood sugar level even in low feed intake (Liukkonen-Anttila, 2001). The decline in cholesterol level with increase in dietary Neem leaf meal is in agreement with the report of Ogbuewu et al. (2008) in a similar work with rabbits. Upadhyay (1990) also reported a decline in blood cholesterol levels of broilers and rats fed Neem leaf meal.

Dietary Neem leaf meal did not significantly (P>0.05) affect the calcium, sodium and potassium levels but significantly (P<0.05) decreased the phosphate level while significantly (P<0.05) and steadily, increasing the chloride level as its level increased. The non-significant increase in serum calcium is an indication that the integrity of the kidney was maintained as reported by Ogbuewu (2008). Serum total protein steadily decreased with increase in dietary Neem leaf meal although the differences were not statistically significant (P>0.05)

Serum albumin and globulin did not show much consistency although at 10% dietary Neem leaf meal inclusion, they became significantly (P<0.05) depressed. Serum albumin and globulin depend on availability of dietary protein. This means that the proteins of the treatments T<sub>0</sub> -T<sub>7.5</sub>, were similarly available to the birds, confirming the earlier observation by Hoffenberg et al. (1966). Urea level declined with Neem leaf meal but creatinine was not affected by the treatments (P>0.05), an indication that the proteins in the diets were effectively utilized. Alanine transaminase (ALT), alkaline phosphatase (ALP) and aspartate transaminase (AST) were depressed as the level of dietary Neem leaf meal increased, indicating no toxic effect within the liver parenchyma of the birds.

Table 4 - Effects of dietary Neem leaf meal on serum biochemical indices of starter broilers						
Indices	Dietary levels of Neem leaf meal					SEM
	T <sub>0</sub>	T <sub>2.5</sub>	T <sub>5.0</sub>	T <sub>7.5</sub>	T <sub>10</sub>	
Glucose (mg/dl)	160.50 <sup>a</sup>	190.47 <sup>ba</sup>	224.73 <sup>b</sup>	195.23 <sup>b</sup>	272.37 <sup>c</sup>	13.22
Calcium (mg/dl)	9.9 <sup>a</sup>	9.57 <sup>a</sup>	10.60 <sup>a</sup>	10.10 <sup>a</sup>	9.80 <sup>a</sup>	0.31
Inorg. Phosphate (mg/dl)	5.93 <sup>a</sup>	4.93 <sup>bc</sup>	5.22 <sup>b</sup>	4.87 <sup>c</sup>	4.17 <sup>c</sup>	0.23
Cholesterol (mg/dl)	144.83 <sup>a</sup>	151.87 <sup>a</sup>	51.87 <sup>c</sup>	77.80 <sup>bc</sup>	85.17 <sup>b</sup>	1.36
Protein (g/dl)	3.20 <sup>a</sup>	3.03 <sup>a</sup>	3.03 <sup>a</sup>	2.90 <sup>a</sup>	2.76 <sup>a</sup>	1.67
Albumin (g/dl)	1.23 <sup>a</sup>	1.37 <sup>ab</sup>	1.47 <sup>a</sup>	1.43 <sup>a</sup>	1.03 <sup>b</sup>	0.07
Globulin (g/dl)	1.96 <sup>a</sup>	1.67 <sup>b</sup>	1.57 <sup>bc</sup>	1.47 <sup>c</sup>	1.73 <sup>a</sup>	0.09
Urea (mg/dl)	30.53 <sup>a</sup>	29.27 <sup>a</sup>	28.03 <sup>a</sup>	27.27 <sup>ab</sup>	27.03 <sup>b</sup>	1.23
Creatinine (mg/dl)	0.90 <sup>a</sup>	0.90 <sup>a</sup>	0.09 <sup>a</sup>	0.09 <sup>a</sup>	0.09 <sup>a</sup>	0.05
Sodium (Mmol/L)	147.93 <sup>a</sup>	153.37 <sup>ab</sup>	158.37 <sup>a</sup>	156.70 <sup>a</sup>	148.53 <sup>a</sup>	1.77
Potassium (Mmol/L)	3.50 <sup>a</sup>	3.20 <sup>a</sup>	3.80 <sup>a</sup>	5.07 <sup>b</sup>	3.87 <sup>a</sup>	0.37
Chloride (Mmol/L)	79.07 <sup>a</sup>	69.60 <sup>a</sup>	78.20 <sup>a</sup>	81.03 <sup>a</sup>	103.10 <sup>b</sup>	3.49
Bicarbonate (Mmol/L)	16.67 <sup>a</sup>	17.17 <sup>a</sup>	22.03 <sup>b</sup>	19.03 <sup>b</sup>	17.17 <sup>a</sup>	0.74
Tot. Billirubin (mg/dl)	0.70 <sup>a</sup>	0.50 <sup>b</sup>	0.37 <sup>b</sup>	0.43 <sup>b</sup>	0.70 <sup>a</sup>	0.51
Conjugated billirubin (mg/dl)	0.37 <sup>a</sup>	0.23 <sup>b</sup>	0.20 <sup>b</sup>	0.20 <sup>b</sup>	0.40 <sup>a</sup>	0.05
ALP (iu/l)	419.77 <sup>a</sup>	395.93 <sup>a</sup>	396.50 <sup>a</sup>	340.77 <sup>b</sup>	311.33 <sup>b</sup>	11.13
ALT (iu/l)	23.33 <sup>a</sup>	19.00 <sup>b</sup>	15.67 <sup>bc</sup>	12.67 <sup>c</sup>	12.00 <sup>c</sup>	1.26
AST (iu/l)	53.33 <sup>a</sup>	43.00 <sup>b</sup>	39.00 <sup>b</sup>	32.00 <sup>c</sup>	18.00 <sup>d</sup>	3.64

<sup>abc</sup> Means within a row with different superscripts are significantly different (P<0.05)

### CONCLUSION

We therefore, concluded that Neem leaf meal can be included in the diets of young broiler chicks up to 10% without any deleterious effects on their haematological and serum biochemical constituents. It reduces blood cholesterol and tends to maintain the integrities of both the kidney and liver.

### REFERENCES

- Aiello SE and Mays M (1998). The Merck-Veterinary Manual, 8<sup>th</sup> edition. Merck and company.
- Akpan MJ (2007). Effects of neem (*Azadirachta indica*) leaf extract on productive performance, egg quality, haematological indices and coccidial load of laying hens. Msc. Thesis, University of uyo, uyo- Nigeria. pp 59-60.
- AOAC (1995). Association of Official Analytical Chemists. Official Methods of Analysis, 7<sup>th</sup> Edition. Washington D.C.

- Babatunde GM, Pond WO, Krook L, Dvan L, Walker ER and Chapman D (1987). Effect of dietary safflower oil or hydrogenated coconut oil on growth rate and on swine blood and tissue components of pigs fed fat-free diets. *Journal of Nutrition*, 92: 1903.
- Church JP, Young JT, Kebau CW, Kebay JC and Ken WW (1984). Relationships among dietary constituents and specific serum clinical components of subjects eating self selected diets. *The American Journal of Clinical Nutrition*, 40: 1338 – 1344.
- Chakraborty T, Verotta L and Podder G (1989). Evaluation of *A. indica* leaf extract for hypoglycaemic activity in rats. *Phytotherapeutic Research*, 3: 30 – 32.
- Dacie JV and Lewis SM (1991). *Practical haematology 7<sup>th</sup> (ed.) ELBS with Churchill living tone*, England pp 37-85.
- Doumas BT (1975). Standards for total Serum Protein assay. *Clinical Chemistry*, 21: 1159.
- Doumas BT and Biggs HG (1972). Determination of serum albumin in standard methods of clinical chemistry. vol.7. (ed.) Cooper, G.R. Academic press New York pp: 175.
- Edozien JC and Switzer B.R (1977). Effects of dietary protein, fat and energy on blood haemoglobin and haematocrit in rat. *Journal of Nutrition*, 107: 1016 – 1021.
- Esonu BO, Emenalom OO, Udedibie ABI, Herbert U, Ekpor CF, Okoli IC and Iheukwumere FC (2001). Performance and blood chemistry of weaner pigs fed raw *Mucuna* bean (velvet) meal. *Tropical Animal Production Investigation*, 4: 49 – 54.
- Esonu BO, Iheukwumere FC, Emenalom OO, Uchegbu MC and Etuk EB (2002). Performance, nutrient utilization and organ characteristics of broilers fed *Microdesmis puberula* leaf meal. *Livestock Research for Rural Development*, 14(16)146.
- Esonu BO, Opara MN, Okoli IC, Obikaonu HO, Udedibie C and Iheshiulor OOM (2006). Physiological Response of Laying Birds to Neem (*Azadirachta indica*) Leaf Meal-Based Diets: Body Weight ,Organ Characteristics and Haematology. *Online Journal of Health and Allied Sciences*, 2:4, www.ojhas.org/issue 18/2006-2-4.htm
- Franson RD (1974). *Anatomy and Physiology of Farm Animals*. Lea & febiger, Philadelphia. 494p.
- Hoffenberg R, Black E and Black JF (1966). Serum Metabolites. *The Journal of Clinical Investigation*, 45: 143-150.
- Henry R, Cannon DC and Winkelman JW (1974). *Clinical chemistry principles and techic* Hagerstown, M. D. Harper, A.H and Row London, pp 543.
- Iyayi EA and Tewe OO (1998). Serum total protein, urea and creatinine levels as indices of quality cassava diets for pigs. *Tropical Veterinary*, 16: 57 – 67.
- Liukkonen-Anttila J (2001). Nutritional and genetic adaptation of gallitorns birds: Implications for hand rearings and resticking. *Acanic Dissertation*, Faculty of Science, University of Oulu, Oulu Yilopisto, Finland. Retrieved September 17, 2007 from <http://herkulesoulu.fi/isbn951425990index.html>
- Okeudo, N. J., Okoli, I. C. and Igwe, G. O. F. (2003). Haematological characteristics of ducks (*Cairina moschata*) of South Eastern Nigeria. *Tropicultura*, 21(2): 61 – 65.
- Ogbuiew IP, Okoli IC and Iloeje MU (2008). Serum biochemical evaluation and organ weight characteristics of buck rabbits fed graded levels of neem (*Azadirachta indica*) leaf meal diets. *Vet online international*. p 3 – 10.
- Nworgu FC, Ogungbenro SA and Solesi KS (2007). Performance and some Blood Chemistry indices of Broiler Chicken served fluted pumpkin (*Telferia occidentalis*) leaves Extract supplement. *American-Eurasian Journal of Agricultural & Environmental Science*, 2(1) 90-98.
- Okorie, K. C. (2006). Evaluation of leaf meals of *Pentaclethra macrophylla*, *Jacaranda mimosifolia* and *Mucuna pruriens* as feed ingredients in poultry diets. PhD Thesis, Federal University of Technology, Owerri – Nigeria. p. 67.
- Sansbury D (1980). *Poultry health and management, chicken, Ducks and Turkeys*. Pp21, 25.
- Steel RGD and Torrie JH (1980). *Principles and procedures of statistics*, New York, McGrae Hill, pp 137-269
- Udedibie, A.B.I. (1987). Comparative evaluation of leaf of paw paw (*C. papaya*), Jackbean (*C. ensiformis*), swordbean (*C. gladiata*) and pigeon pea (*C. cajan*) as feed ingredients and yolk coloring agents in laying diets. *Nigeria Journal of Animal Production*, 14: 61-66.
- Udedibie ABI and Opara CC (1996). Response of growing broilers and leaf meal from *Alchonea cordifolia* Anim. *Food Science Technology*, 71: 157-164.
- Upadhyay C (1990). The medicinal properties of neem (*Azadirachta indica*) tree. In: *Animal pharmacology 2<sup>nd</sup> edition*. Longman England.
- Varley H, Gowshock AH and Bell M (1980). *Determination of Serum Urea using Biochemistry*. 5<sup>th</sup> Edition William Heineman Medical Books, Ltd., London.
- Vohra P, Hjenrick RB, Wilson WC and Scopes TD (1972). The Use of ipil-ipil (*Leucaena leucocephala*) in the diets of laying chickens and laying quails. *The Philippine Agriculturist*, 56: 104-133.
- WHO. (1980). *Manual of Basic Techniques for a Health Laboratory*. World Health Organization, Geneva, pp. 360 – 406.